

**ABOUT THE TEST** FoundationOne®Heme is a comprehensive genomic profiling test designed to identify genomic alterations within hundreds of cancer-related genes in hematologic malignancies, sarcomas, and pediatric cancers.

**PATIENT**

**DISEASE** Acute myeloid leukemia (AML) (NOS)  
**NAME** Not Given  
**DATE OF BIRTH** Not Given  
**SEX** Female  
**MEDICAL RECORD #** Not Given

**PHYSICIAN**

**ORDERING PHYSICIAN** Not Given  
**MEDICAL FACILITY** Not Given  
**ADDITIONAL RECIPIENT** Not Given  
**MEDICAL FACILITY ID** Not Given  
**PATHOLOGIST** Not Given

**SPECIMEN**

**SPECIMEN SITE** Not Given  
**SPECIMEN ID** Not Given  
**SPECIMEN TYPE** Not Given  
**DATE OF COLLECTION** Not Given  
**SPECIMEN RECEIVED** Not Given

**Biomarker Findings**

**Microsatellite status - MS-Stable**  
**Tumor Mutational Burden - TMB-Low (2 Muts/Mb)**

**Genomic Findings**

*For a complete list of the genes assayed, please refer to the Appendix.*

**KIT D816V**  
**NRAS Q61H**  
**MYH11 CBF3-MYH11 fusion**

**5 Therapies with Clinical Benefit**      **10 Clinical Trials**  
**2 Therapies with Lack of Response**

**BIOMARKER FINDINGS**

**Microsatellite status - MS-Stable**

**Tumor Mutational Burden - TMB-Low (2 Muts/Mb)**

**THERAPIES WITH CLINICAL BENEFIT (IN PATIENT'S TUMOR TYPE)**

No therapies or clinical trials. see Biomarker Findings section

**THERAPIES WITH CLINICAL BENEFIT (IN OTHER TUMOR TYPE)**

No therapies or clinical trials. see Biomarker Findings section

**GENOMIC FINDINGS**

**KIT - D816V**

**7 Trials** see p. 16

**NRAS - Q61H**

**3 Trials** see p. 17

**THERAPIES WITH CLINICAL BENEFIT (IN PATIENT'S TUMOR TYPE)**

None

**THERAPIES WITH CLINICAL BENEFIT (IN OTHER TUMOR TYPE)**

Dasatinib  
Nilotinib  
Ponatinib  
▲ Imatinib<sup>1</sup>  
▲ Sunitinib<sup>1</sup>

None

Cobimetinib  
Trametinib

▲ 1. Patient may be resistant to indicated therapy

**GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIALS OPTIONS**

*For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.*

**MYH11 - CBF3-MYH11 fusion** ..... p. 18

**NOTE** Genomic alterations detected may be associated with activity of certain FDA-approved drugs; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type.

TRF# XXXXXXXX

**BIOMARKER FINDINGS**
**BIOMARKER**

## Tumor Mutational Burden

**CATEGORY**

TMB-Low (2 Muts/Mb)

**POTENTIAL TREATMENT STRATEGIES**

On the basis of emerging clinical evidence, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-CTLA-4<sup>148</sup>, anti-PDL1<sup>149,151,152</sup>, and anti-PD-1 therapies<sup>122,142,153</sup>; FDA-approved agents include ipilimumab, atezolizumab, avelumab, durvalumab, pembrolizumab, and nivolumab. In multiple solid tumor types, higher mutational burden has corresponded with response and improved prognosis. Pembrolizumab improved progression-free survival (14.5 vs. 3.4-3.7 months) in patients with non-small cell lung cancer (NSCLC) and higher mutational load (greater than 200 nonsynonymous mutations; hazard ratio = 0.19)<sup>142</sup>. In studies of patients with either NSCLC or colorectal cancer (CRC), patients whose tumors harbor elevated mutational burden reported higher overall response rates to

pembrolizumab<sup>122,142,153</sup>. Anti-PD-1 therapies have achieved clinical benefit for certain patients with high mutational burden, including 3 patients with endometrial adenocarcinoma who reported sustained partial responses following treatment with pembrolizumab<sup>154</sup> or nivolumab<sup>155</sup>, a patient with hypermutant glioblastoma who obtained clinical benefit from pembrolizumab<sup>156</sup>, and two pediatric patients with biallelic mismatch repair deficiency (bMMRD)-associated ultrahypermutant glioblastoma who experienced clinically and radiologically significant responses to ivolumab<sup>157</sup>. In patients with melanoma, mutational load was associated with long-term clinical benefit from ipilimumab<sup>148,158</sup> and anti-PD-1/anti-PD-L1 treatments<sup>151</sup>. For patients with metastatic urothelial carcinoma, those who responded to atezolizumab treatment had a significantly increased mutational load [12.4 mutations (mut) per megabase (Mb)] compared to nonresponders (6.4 mut/Mb)<sup>149</sup>, and mutational load of 16 mut/Mb or higher was associated with significantly longer overall survival<sup>152</sup>.

**FREQUENCY & PROGNOSIS**

Reports of high TMB are generally rare in leukemia<sup>150</sup>. In a study of 92 patients with various hematologic malignancies, elevated TMB levels [ $>10$  mutations per megabase

(mut/Mb)] were not detected in AML (0/5) or ALL (0/1) cases analyzed (Karim et al., 2017; AACR Abstract 3724).

**FINDING SUMMARY**

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma<sup>139,140</sup> and cigarette smoke in lung cancer<sup>141,142</sup>, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes<sup>143,144,145,146,147</sup>, and microsatellite instability (MSI)<sup>143,146,147</sup>. The tumor seen here harbors a low TMB. Compared to patients with tumors harboring higher TMB levels, patients with tumors harboring low TMB levels have experienced lower rates of clinical benefit from treatment with immune checkpoint inhibitors, including anti-CTLA-4 therapy in melanoma<sup>148</sup>, anti-PD-L1 therapy in urothelial carcinoma<sup>149</sup>, and anti-PD-1 therapy in non-small cell lung cancer and colorectal cancer<sup>122,142</sup>.

**BIOMARKER**

## Microsatellite status

**CATEGORY**

MS-Stable

**POTENTIAL TREATMENT STRATEGIES**

On the basis of clinical evidence, microsatellite stable (MSS) tumors are significantly less likely than MSI-high (MSI-H) tumors to respond to anti-PD-1 immune checkpoint inhibitors<sup>119,120,121</sup>, including approved therapies nivolumab and pembrolizumab (Overman et al., 2016; ASCO Abstract 3501)<sup>122</sup>. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%,  $p=0.001$ ) (Ayers et al., ASCO-SITC 2016; Abstract P60). Pembrolizumab therapy resulted in a significantly lower objective response rate (ORR) in MSS colorectal cancer (CRC) compared with MSI-H CRC (0% vs. 40%)<sup>122</sup>. Similarly, a clinical study of nivolumab, alone or in combination with ipilimumab, in patients with CRC reported a

significantly higher response rate in patients with MSI-H tumors than those without (Overman et al., 2016; ASCO Abstract 3501).

**FREQUENCY & PROGNOSIS**

High MSI (MSI-H) is generally rare in hematologic malignancies compared with solid tumors. Moreover, reports of MSI in hematologic malignancies in the literature are conflicting and varied due to substantial heterogeneity, lack of consensus on the markers and methods used for MSI assessment, small sample size in most studies, and possible elimination of MSI positive cells in the bloodstream by immunosurveillance<sup>99</sup>. In studies of acute myeloid leukemia (AML), MSI at any level has been reported at incidences from 6-56%<sup>100,101,102,103,104,105,106,107</sup>; however, contradicting studies reported an absence of MSI in AML (Bonneville et al., 2017; DOI: 10.1200/PO.17.00073)<sup>108</sup>. Similarly, MSI-H has been observed with incidences of 3-32%<sup>102,104,105,107</sup> or reported as absent in AML (Bonneville et al., 2017; DOI: 10.1200/PO.17.00073)<sup>100</sup>. In a large study of 1,394 patients with de novo or therapy-related AML, MSI-H was not observed; however, 4.8% of cases demonstrated instability at one microsatellite locus<sup>109</sup>. In addition, a small number of studies have not found a significant correlation of MSI with

relapsed AML<sup>104</sup>, nor with progression from MDS to AML<sup>110</sup>, and other publications have reported a high incidence (20-32%) of MSI in de novo AML/MDS<sup>105,106,107,111</sup>. In contrast, other studies have reported increased incidences of MSI in relapsed or therapy-related AML/MDS compared to de novo disease<sup>103,107,111,112,113,114,115,116</sup>, and a cell lineage analysis of AML/CML progression found increased MSI associated with relapsed disease after chemotherapy in 3/6 patients<sup>117</sup>. Therefore, the role of MSI in MDS/AML progression and resistance to chemotherapy is unclear. One study has suggested that organ transplant patients are at higher risk of developing AML/MDS as a result of prolonged immunosuppression, and reported all 7 such patients analyzed exhibited MSI, with 6/7 being MSI-H<sup>118</sup>.

**FINDING SUMMARY**

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor<sup>93</sup>. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2<sup>93,94,95</sup>. The tumor

TRF# XXXXXXXX

**BIOMARKER FINDINGS**

seen here is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite

markers 96,97,98. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins 93,95,97,98.

SAMPLE

TRF# XXXXXXXX

**GENOMIC FINDINGS**

**GENE**

**KIT**

**ALTERATION**

**D816V**

**POTENTIAL TREATMENT STRATEGIES**

On the basis of clinical evidence, primarily in GIST, AML and systemic mastocytosis, KIT activating alterations are associated with sensitivity to KIT tyrosine kinase inhibitors including imatinib, sunitinib, sorafenib, dasatinib, nilotinib, pazopanib, regorafenib, ponatinib, midostaurin and avapritinib (Heinrich et al., 2014; ASCO Abstract 10506, Heinrich et al., 2017; ASCO Abstract 11011, DeAngelo et al., 2017; ASH Abstract 2)21,28,29,30,31,32,33,34. However, mutation of residues within the KIT activation-loop (A-loop) has been associated with preclinical and clinical resistance to imatinib (Heinrich et al., 2013; ASCO Abstract 10509)5,6,7,8,9,10,11,12,13,14,15,16,17,35,36 and sunitinib (Heinrich et al., 2013; ASCO Abstract 10509)5,10,14,17,37 in patients with GIST. KIT exon 17 mutations, including at D816 were reported to be sensitive to avapritinib in clinical (Heinrich et al., 2017; ASCO Abstract 11011, DeAngelo et al., 2017;

ASH Abstract 2)21 and preclinical21 studies. Preclinical38,39,40 and limited clinical41,42,43 data indicate that KIT A-loop mutants are sensitive to sorafenib, although preclinical evidence of potential resistance has been reported specifically for the D816V mutation39,44,45. Several Aloop alterations at residues D816, D820, N822, and A829 have exhibited ponatinib sensitivity in preclinical evaluations (Schrock et al., 2013; ASCO Abstract B266)17,46,47,48,49. Limited clinical evidence in melanoma suggests that mTOR inhibitors as a monotherapy or in combination with first-line kinase inhibitors may be a useful strategy to target kinase inhibitor-resistant tumors (Si et al., 2012; ASCO Abstract 8562)50.

**FREQUENCY & PROGNOSIS**

KIT mutations have been reported in 4-17% of acute myeloid leukemia (AML) samples22,23, and in 30% of CBF-AML cases24. KIT expression has been reported 60-80% of patients with AML23. Reports on the prognostic value of KIT mutations in AML have been mixed24,25, but some studies have found that patients with KIT mutations have a poorer prognosis than those without24,26,27.

**FINDING SUMMARY**

KIT (also called c-KIT) encodes a cell surface tyrosine kinase receptor that, upon ligand binding and dimerization, activates the PI3K-AKT and RAS-

MAPK signaling pathways1. KIT aberrations, including point mutations, translocations, amplification, and overexpression, have been associated with various malignancies, and KIT is considered an oncoprotein2. Activating alterations in KIT, or the related gene PDGFRA, are known to underlie inherited predisposition to gastrointestinal stromal tumor (GIST) development3,4. Mutations of KIT within the activation loop, including amino acids C809, D816, D820, N822, Y823, and A829, as also observed here, have been reported to confer preclinical and clinical resistance to imatinib (Heinrich et al., 2013; ASCO Abstract 10509)5,6,7,8,9,10,11,12,13,14,15,16,17 and sunitinib (Heinrich et al., 2013; ASCO Abstract 10509)5,10,17,18 in GIST. KIT D816G has also been reported as an emergent mutation conferring resistance to crizotinib in a patient with ROS1-rearranged non-small cell lung carcinoma (NSCLC)19. In a rare case, a partial response to imatinib was observed in a patient with a KIT activation loop mutation20. KIT exon 17 mutations, including at D816 were reported to be sensitive to avapritinib in clinical (Heinrich et al., 2017; ASCO Abstract 11011, DeAngelo et al., 2017; ASH Abstract 2)21 and preclinical21 studies.

**GENE**

**MYH11**

**ALTERATION**

**CBFB-MYH11 fusion**

**POTENTIAL TREATMENT STRATEGIES**

There are no targeted therapies are available to address genomic alterations in CBFB.

**FREQUENCY & PROGNOSIS**

CBFB-MYH11 fusions are present in 5% of acute myeloid leukemia (AML) cases124. The presence of either the CBFB-MYH11 fusion or RUNX1-RUNX1T1 (AML-ETO) fusion defines a subtype of

AML (CBF-AML) that is associated with favorable prognosis124,130,131,132,133. No difference in prognosis has been noted between the different variants of CBFB-MYH11 fusions observed thus far134,135. However, high copy numbers of the CBFB-MYH11 fusion after induction therapy has been associated with higher relapse rate in CBF-AML versus patients with low copy numbers136,137. CBFBMYH11 fusion is thought to be insufficient for leukemic transformation by itself, and is frequently accompanied by other rearrangements or alterations, including alterations to the RAS pathway in up to 90% of samples71,124,127,138.

**FINDING SUMMARY**

CBFB encodes the regulatory beta subunit of core binding factor, CBF-beta, that complexes with CBF-alpha (RUNX1, 2, or 3) to regulate transcription of

genes required for normal hematopoiesis and osteogenesis123. CBFB-MYH11 fusions commonly result from inv(16)(p13.1q22) or t(16;16)(p13.1;q22)124 and share the N-terminus (most commonly exons 1-5, aa 1-165) of CBFB and Cterminal dimerization domain of smooth muscle myosin heavy chain125,126. CBFB-MYH11 fusion interferes with the transcriptional regulation mediated by RUNX1 (and other RUNX family members), leading to impaired hematopoietic cell differentiation and predisposition to leukemic transformation127,128; however, additional, RUNX1-independent mechanisms have also been proposed129.

TRF# XXXXXXXX

**GENOMIC FINDINGS**

**GENE**  
**NRAS**

**ALTERATION**  
**Q61H**

mutation predicts sensitivity to the PI3K-alpha-specific inhibitor BYL71980. The reovirus Reolysin targets cells with activated RAS signaling<sup>81,82,83</sup> and is in clinical trials in patients with some tumor types. Reolysin has demonstrated mixed clinical efficacy, with the highest rate of response reported for patients with head and neck cancer<sup>84,85,86,87,88,89,90,91,92</sup>.

kinase inhibitors (TKIs) in AML cell lines may be driven by activating mutations in NRAS, leading to upregulation of AKT and MAPK signaling; combination of FLT3 TKIs with inhibitors of PI3K or MEK may circumvent this resistance<sup>72</sup>.

**FINDING SUMMARY**

NRAS encodes a member of the RAS family of small GTPases that mediate transduction of growth signals. Activation of RAS signaling causes cell growth, differentiation, and survival by activating the RAF-MAPK-ERK, PI3K, and other pathways<sup>51</sup>. NRAS alterations affecting amino acids G12, G13, G60, and Q61 as well as mutations I24N, T50I, and A146T have been characterized to be activating and oncogenic<sup>51,52,53,54,55,56,57,58,59,60,61,62,63,64,65</sup>.

**POTENTIAL TREATMENT STRATEGIES**

Preclinical evidence suggests that NRAS activation may predict sensitivity to MEK inhibitors, such as cobimetinib and trametinib<sup>51,73,74,75,76</sup>. Clinical responses to MEK inhibitor-based therapy regimens have been observed in patients with NRAS mutation in certain tumor types (Heuck et al., 2014; ASH Abstract 4775)<sup>77,78,79</sup>. Preclinical data in cancer cell lines indicates that NRAS

**FREQUENCY & PROGNOSIS**

NRAS mutation has been reported in 8-25%<sup>22,66</sup> of acute myeloid leukemia (AML) cases. Studies have reported that RAS mutation (NRAS or KRAS) had no influence on clinical outcome in pediatric patients with AML, in patients under 60 years old with AML, in patients with secondary AML, or in patients with AML harboring CBFβ-MYH11 fusions<sup>67,68,69,70,71</sup>. A preclinical study suggested that acquired resistance to FLT3 tyrosine



TRF# XXXXXXXX

**THERAPIES WITH CLINICAL BENEFIT**
**IN OTHER TUMOR TYPE**

## Cobimetinib

*Assay findings associations*
**NRAS**  
 Q61H

### AREAS OF THERAPEUTIC USE

Cobimetinib is a MEK inhibitor that is FDA approved in combination with vemurafenib for the treatment of unresectable or metastatic melanoma with BRAF V600E or V600K mutations.

### GENE ASSOCIATION

On the basis of a case study in an NRAS-mutant chronic myelomonocytic leukemia (CMML)<sup>208</sup>, NRAS amplification or activating mutations may predict sensitivity to MEK inhibitors such as cobimetinib. Significant clinical responses to various other MEK inhibitors have been documented in NRAS-mutant melanoma<sup>77,209,210,211</sup>.

### SUPPORTING DATA

Cobimetinib has been investigated primarily in the context of BRAF V600-mutant melanoma. A Phase 3 study with 495 patients treated either with the BRAF inhibitor vemurafenib plus cobimetinib or vemurafenib alone

reported a 68-70% overall response rate, 9.9-12.3 months progression-free survival, and a lower rate of cutaneous squamous cell carcinoma in the combination group; disease progression did not correlate with concurrent alterations in the RAS pathway (Larkin et al., 2015; ASCO Abstract 9006)<sup>212</sup>. In a Phase 1b study, vemurafenib combined with cobimetinib achieved an objective response rate of 87% for patients with BRAF V600-mutant melanoma who had not previously received a BRAF inhibitor<sup>213</sup>. One study reported near-complete response to vemurafenib in a patient with BRAF V600K-mutant melanoma who subsequently developed chronic myelomonocytic leukemia (CMML) with NRAS G12R mutation, and concurrent cobimetinib treatment led to suppression of CMML<sup>208</sup>. In a Phase 1b study, out of 47 patients treated with cobimetinib and the AKT inhibitor ipatasertib, 3 patients with KRAS-mutant ovarian, mesonephric cervical, or endometrial carcinoma had a partial response, with prolonged stable disease lasting for >6 months (Bendell et al., 2014; AACR Abstract CT328).

## Dasatinib

*Assay findings associations*
**KIT**  
 D816V

### AREAS OF THERAPEUTIC USE

Dasatinib is a kinase inhibitor that targets the BCR-ABL fusion protein, SRC family kinase receptors, KIT, EPHA2, and PDGFR-beta. It is FDA approved for the treatment of certain subtypes of Philadelphia chromosome-positive (Ph+) chronic myeloid leukemia (CML) and Ph+ acute lymphoblastic leukemia (ALL).

### GENE ASSOCIATION

On the basis of clinical evidence in melanoma (Kalinsky et al., 2016; ASCO 2016 Abstract 9501)<sup>159</sup> and preclinical evidence in other cancer types<sup>160,161</sup>, activating KIT alterations may confer sensitivity to dasatinib. Preclinical and limited clinical data indicate that secondary

imatinibresistant mutations, including V654A and A-loop alterations at residues D816, D820, N822, and A829, remain sensitive to dasatinib<sup>38,161,162,163,164,165</sup>.

### SUPPORTING DATA

A case study has reported long term remission in an acute myeloid leukemia (AML) patient with a KIT D816V mutation treated with a combination of chemotherapy and dasatinib<sup>165</sup>. Preclinical studies have reported mixed responses to dasatinib in small studies of unselected AML patients<sup>166</sup>. Inhibition of cell lines expressing KIT D816V following treatment with dasatinib has been achieved, but the levels of drug required exceed those that are likely to be achievable in patients<sup>167,168</sup>.



TRF# XXXXXXXX

**THERAPIES WITH CLINICAL BENEFIT**    **IN OTHER TUMOR TYPE**

**Nilotinib**

*Assay findings associations*

**KIT**  
D816V

**AREAS OF THERAPEUTIC USE**

Nilotinib targets tyrosine kinases such as ABL (including BCR-ABL), PDGFRs, KIT, CSF1R, DDR1, and DDR2. It is FDA approved to treat newly diagnosed pediatric or adult patients with Philadelphia chromosome-positive (Ph+) chronic myeloid leukemia (CML) in chronic phase, adults with Ph+ CML in chronic or accelerated phase with resistance or intolerance to prior therapy including imatinib, and pediatric patients with Ph+ CML in chronic phase with resistance or intolerance to prior tyrosine-kinase inhibitor therapy.

**GENE ASSOCIATION**

On the basis of strong clinical<sup>186,187,188,189</sup> and preclinical<sup>190,191</sup> data in multiple tumor types, KIT amplification or activation may confer sensitivity to nilotinib. Additionally, imatinib-resistant KIT mutations such as V654A and A-loop alterations at residues D816, D820, N822, and A829 maintain sensitivity to nilotinib as

demonstrated in multiple clinical and preclinical studies<sup>38,162,163,188,192,193,194</sup>.

**SUPPORTING DATA**

A Phase 2 study of nilotinib in combination with daunorubicin and cytarabine in patients with newly diagnosed acute myeloid leukemia (AML) and KIT expression but no KIT mutations reported a complete response (CR) in 80% (12/15) of evaluable patients, and a low relapse rate of 17% (2/12) in patients who achieved CR (Al-Kali et al., 2015; ASH Annual Meeting Abstract 3808). A Phase 1 study of nilotinib in combination with re-induction chemotherapy in patients with relapsed and refractory KIT-positive AML reported a high CR rate (83%, 10/12 patients overall) but noted that the concentrations needed to inhibit the KIT pathway were generally higher than clinically achievable (Brandwein et al., 2013; ASH Annual Meeting Abstract 3961).

**Ponatinib**

*Assay findings associations*

**KIT**  
D816V

**AREAS OF THERAPEUTIC USE**

Ponatinib is a multi-kinase inhibitor targeting BCR-ABL, RET, KIT, FLT-3, PDGFRs, VEGFRs, FGFRs, and other tyrosine kinases. Ponatinib was approved by the FDA for use in advanced, T315I-mutated chronic myeloid leukemia (CML) and Philadelphia chromosome-positive acute lymphoblastic leukemia (Ph+ ALL), as well as for CML and Ph+ ALL patients for whom no other tyrosine kinase inhibitor is indicated.

**GENE ASSOCIATION**

KIT exon 9 and 11 activating mutations have demonstrated sensitivity to ponatinib in several preclinical cell models (Schrock et al., 2013; ASCO Abstract B266, Garner et al., 2013; AACR Abstract 3394)<sup>46,47,48,49</sup>, with clinical efficacy demonstrated in patients with GIST exhibiting exon 11 mutations (Heinrich et al., 2014; ASCO Abstract 1050)<sup>17</sup> and

imatinib resistance (Heinrich et al., 2014; ASCO Abstract 10506). Secondary imatinib-resistant mutations, including T670I (gatekeeper) and Aloop alterations at residues D816, D820, N822, and A829, have exhibited ponatinib sensitivity in preclinical evaluations (Schrock et al., 2013; ASCO Abstract B266)<sup>17,46,47,48,49</sup>.

**SUPPORTING DATA**

Clinically, ponatinib has been most extensively studied in patients with BCR-ABL-positive hematological malignancies. Ponatinib has shown efficacy in preclinical models of endometrial, bladder, gastric, breast, lung, colon, and medullary thyroid carcinomas (Gozgit et al., 2013; AACR Abstract 2084)<sup>195</sup>. A Phase 1 study of ponatinib in patients with acute myeloid leukemia (AML), all of whom had FLT3 alterations, reported 3/12 response rate, with two complete responses and one partial response<sup>196</sup>.

TRF# XXXXXXXX

**THERAPIES WITH CLINICAL BENEFIT**
**IN OTHER TUMOR TYPE**

## Trametinib

*Assay findings associations*
**NRAS**  
 Q61H

### AREAS OF THERAPEUTIC USE

Trametinib is a MEK inhibitor that is FDA approved as a monotherapy and in combination with dabrafenib to treat patients with unresectable or metastatic melanoma with BRAF V600E or V600K mutations, as well as in combination with dabrafenib as adjuvant treatment for completely resected advanced BRAF V600E- or V600K-positive melanoma. It is also approved in combination with dabrafenib to treat patients with metastatic non-small cell lung cancer (NSCLC) with a BRAF V600E mutation and to treat patients with BRAF V600E-positive anaplastic thyroid cancer (ATC) who lack satisfactory locoregional treatment options.

### GENE ASSOCIATION

Activating mutations in NRAS may result in activation of downstream pathways, including the MAPK pathway, and may therefore predict sensitivity to inhibitors of MAPK pathway components, including the MEK inhibitor trametinib. A patient with atypical chronic myeloid leukemia harboring an NRAS mutation exhibited a durable, near-complete response on trametinib treatment<sup>79</sup>. Significant clinical responses to various other MEK inhibitors have been documented in NRAS-mutant melanoma<sup>77,209,210,211</sup>.

### SUPPORTING DATA


A Phase 1/2 study evaluated trametinib for the treatment of relapsed or refractory myeloid malignancies<sup>214</sup>. Patients with KRAS- or NRAS-mutated acute myeloid leukemia (AML) or myelodysplastic syndrome achieved an overall response rate (ORR) of 20% (10/50), including 6 complete remissions (CRs), and a median overall survival (OS) of 4.9 months. Patients with KRAS- or NRAS-mutated chronic myelomonocytic leukemia had an ORR of 27% (3/11), including 3 CRs, and a median OS of 14.5 months. In contrast, the study reported an ORR of 3% (1/30) and a median OS of 3.0 months for patients with wild-type or unknown RAS status<sup>214</sup>. Retrospective genomic analysis of RAS-mutated cases suggested that mutations in epigenetic regulators (e.g., MLL2, SETD2, TET2, IDH1/2) were more frequent among nonresponders than responders (70% vs. 33%) (Johnson et al., 2015; ASH Abstract 1386). Preclinical data support the sensitivity of RAS-mutated AML to MEK inhibitors, including trametinib<sup>73,215</sup>. A patient with NRAS-mutated atypical chronic myeloid leukemia (CML) experienced an exceptional hematologic response and disease control for at least 14 months on trametinib therapy<sup>79</sup>. Another patient with CML and concomitant melanoma reported a complete hematological response to treatment with dabrafenib and trametinib<sup>216</sup>.



TRF# XXXXXXXX

**THERAPIES ASSOCIATED WITH LACK OF RESPONSE**    **IN OTHER TUMOR TYPE**

**Imatinib**

 Patient may be resistant to Imatinib

Assay findings associations

**KIT**  
D816V

**APPROVED INDICATIONS**

Imatinib targets the BCR-ABL fusion protein, PDGFR, and KIT. It is FDA approved for the treatment of KIT-positive gastrointestinal stromal tumors (GIST), Ph+ chronic myeloid leukemia (CML) and acute lymphoblastic leukemia (ALL), myelodysplastic syndrome/myeloproliferative syndrome (MDS/MPS), aggressive systemic mastocytosis without a D816V KIT mutation, hypereosinophilic syndrome and/or chronic eosinophilic leukemia, and dermatofibrosarcoma protuberans.


**GENE ASSOCIATION**

KIT activating mutations or amplification may confer sensitivity to tyrosine kinase inhibitors such as imatinib<sup>20,169,170,171,172,173,174,175,176,177</sup>. However, mutation of residues within or adjacent to the KIT activation loop, as observed here, have demonstrated resistance to imatinib in patients with GIST<sup>9,11,13,35,36,178,179,180,181,182</sup> (Liegl B, et al. 2008; 18623623).

**SUPPORTING DATA**

A Phase 2 study of imatinib in patients with KIT-positive acute myeloid leukemia (AML) reported responses in 5 out of 21 cases, including 2 complete hematologic remissions, 1 patient with no evidence of leukemia, and 2 sustained minor responses<sup>175</sup>. However, other studies reported limited efficacy of imatinib in KIT-positive AML, with a Phase 2 study reporting clinical response consisting of hematologic improvement in 1 out of 36 patients<sup>183</sup> and another study reporting no response in any of 10 included patients with AML or 8 with myelodysplastic syndrome (MDS)<sup>184</sup>. A Phase 1/2 study of imatinib with mitoxantrone, etoposide, and cytarabine in KIT-positive AML reported a complete response rate of 62% (13/21)<sup>169</sup>. A case study reported a complete molecular response to single-agent imatinib maintenance lasting more than 6 months in a patient with BCR-ABL-positive AML<sup>185</sup>.

**Sunitinib**

 Patient may be resistant to Sunitinib

Assay findings associations

**KIT**  
D816V

**APPROVED INDICATIONS**

Sunitinib is a small-molecule tyrosine kinase inhibitor that targets PDGFRs, VEGFRs, KIT, FLT3, CSF-1R, and RET. It is FDA approved for the treatment of advanced or metastatic pancreatic neuroendocrine tumors, gastrointestinal stromal tumors (GISTs) in patients who have progressed on or are intolerant to imatinib, and advanced renal cell carcinoma (RCC) as well as for the adjuvant treatment of patients at high risk of recurrent RCC after nephrectomy.

**GENE ASSOCIATION**

On the basis of extensive clinical and preclinical data in multiple tumor types<sup>10,197,198,199,200,201,202,203</sup>, KIT amplification or activating mutation may predict sensitivity to sunitinib. However, mutation of residues within the KIT

activation loop (A-loop) is predicted to confer resistance to sunitinib based on extensive clinical data in GIST<sup>5,10,14,37</sup>.

**SUPPORTING DATA**

Sunitinib has been shown to inhibit activated FLT3 and the proliferation of cells with FLT3-activating mutations in preclinical studies of acute myeloid leukemia (AML)<sup>204,205</sup>. In one clinical study, 4/4 AML patients with activating FLT3 mutations exhibited morphologic or partial responses to sunitinib<sup>206</sup>. A case study reported complete though short-term hematologic responses for a patient with an eosinophilia-associated myeloid neoplasm and ETV6-FLT3 fusion, after sequential sunitinib and sorafenib therapeutic regimens<sup>207</sup>.

**NOTE** Genomic alterations detected may be associated with activity of certain FDA approved drugs, however the agents listed in this report may have little or no evidence in the patient's tumor type.

TRF# XXXXXXXX

**CLINICAL TRIALS**

**IMPORTANT** Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually

updated and should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that

may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://www.foundationmedicine.com/genomic-testing#support-services). Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

**GENE**  
**KIT**
**ALTERATION**  
**D816V**
**RATIONALE**

KIT amplification or activating mutations may predict sensitivity to small molecule tyrosine kinase inhibitors. Also, because KIT activation leads to activation of the PI3K-AKT-mTOR pathway, PI3K and mTOR inhibitors may be relevant in a tumor with KIT activation. However, mutation of the KIT activation loop, as seen here, is associated with secondary resistance to imatinib and sunitinib; therefore, imatinib and

sunitinib may not be effective in this case. Examples of clinical trials that may be appropriate for this patient are listed below. These trials were identified through a search of the trial website [clinicaltrials.gov](https://www.clinicaltrials.gov) using keyword terms such as "KIT", "PI3K", "mTOR", "sorafenib", "dasatinib", "nilotinib", "PKC412", "ponatinib", "everolimus", "temsirolimus", "AML", and/or "leukemia".

**NCT01643603**
**PHASE 1 / 2**

Phase I/II Study of Dasatinib in Recipients of Allogeneic Stem Cell Transplantation for Hematologic Malignancies.

**TARGETS**  
**ABL, SRC, DDR2, KIT, PDGFRs**
**LOCATIONS:** Michigan

**NCT01806571**
**PHASE 2**

A Phase II Study of Combination Daunorubicin and Cytarabine (Ara-c) and Nilotinib (Tasigna) (DATA) in Patients Newly Diagnosed With Acute Myeloid Leukemia and KIT Overexpression

**TARGETS**  
**ABL, KIT, PDGFRs**
**LOCATIONS:** Arizona, Minnesota

**NCT01552434**
**PHASE 1**

A Phase I Trial of Bevacizumab, Temsirolimus Alone and in Combination With Valproic Acid or Cetuximab in Patients With Advanced Malignancy and Other Indications

**TARGETS**  
**HDAC, EGFR, VEGFA, mTOR**
**LOCATIONS:** Texas

**NCT01582191**
**PHASE 1**

A Phase 1 Trial of Vandetanib (a Multi-kinase Inhibitor of EGFR, VEGFR and RET Inhibitor) in Combination With Everolimus (an mTOR Inhibitor) in Advanced Cancer

**TARGETS**  
**EGFR, RET, SRC, VEGFRs, mTOR**
**LOCATIONS:** Texas

**NCT01620216**
**PHASE 2**

A Phase II Pilot Study of Kinase Inhibition in Relapsed/Refractory Acute Leukemias: Using a Comprehensive in Vitro Kinase Inhibitor Panel to Select Individualized, Targeted Therapies.

**TARGETS**  
**CSFR1, FLT3, KIT, PDGFRs, RET, VEGFRs, BCR-ABL, BTK, EPHA2, LYN, SRC, DDR1, BRAF, CRAF, EPH, TIE2, FGFRs**
**LOCATIONS:** Texas

TRF# XXXXXXXX

**CLINICAL TRIALS**

**NCT02779283**

**PHASE 1**

A Phase Ib Feasibility Study of Personalized Kinase Inhibitor Therapy Combined With Induction Chemotherapy in Acute Myeloid Leukemia in Patients Who Exhibit In Vitro Kinase Inhibitor Sensitivity

**TARGETS**  
ABL, FGFRs, RAFs, RET, SRC, CSF1R,  
DDR2, FLT3, KIT, PDGFRs, VEGFRs

**LOCATIONS:** Oregon

**NCT02013648**

**PHASE 3**

Randomized Phase III Study of Intensive Chemotherapy With or Without Dasatinib (Sprycel™) in Adult Patients With Newly Diagnosed Core-Binding Factor Acute Myeloid Leukemia (CBF-AML)

**TARGETS**  
ABL1, SRC, KIT, EPHA2, PDGFRs, LYN

**LOCATIONS:** Multiple ex-US locations

SAMPLE

TRF# XXXXXXXX

**CLINICAL TRIALS**

GENE  
**NRAS**

ALTERATION  
Q61H

**RATIONALE**

Activating mutations in NRAS leads to activation of the RAF-MEK-ERK, PI3K, and other pathways, and may predict sensitivity to inhibitors of these downstream pathways. Examples of clinical trials that may be appropriate for this patient are listed below. These trials were identified through a search

of the trial website [clinicaltrials.gov](http://clinicaltrials.gov) using keyword terms such as "NRAS", "MEK", "PI3K", "MEK162", "BKM120", "selumetinib", "trametinib", "cobimetinib", "binimetinib", "refametinib", "E6201", "reolysin", "AML", "leukemia", and/or "advanced cancer".

<p><b>NCT02670044</b></p> <p>A Phase IB/II Multi-Arm Study With Venetoclax in Combination With Cobimetinib and Venetoclax in Combination With Idasanutlin in Patients Aged &gt;= 60 Years With Relapsed or Refractory Acute Myeloid Leukemia Who Are Not Eligible for Cytotoxic Therapy</p>	<p><b>PHASE 1 / 2</b></p> <p><b>TARGETS</b> BCL2, MDM2, MEK</p>
<p><b>LOCATIONS:</b> Colorado, Roma (Italy), Marseille (France), Toronto (Canada), Massachusetts, Bobigny (France), Bologna (Italy), North Carolina, Texas, Montreal (Canada), Pesaro (Italy), California, New York, Edmonton (Canada), Pessac (France)</p>	
<p><b>NCT02089230</b></p> <p>Phase I/II Trial of MEK Inhibitor MEK162 in Patients With Relapsed and or Refractory Acute Myeloid Leukemia and Patients With Poor Prognosis Acute Myeloid Leukemia Not Suitable for or Unwilling to Receive Standard Therapy</p>	<p><b>PHASE 1 / 2</b></p> <p><b>TARGETS</b> MEK</p>
<p><b>LOCATIONS:</b> Texas</p>	
<p><b>NCT02551718</b></p> <p>Individualized Treatment for Relapsed/Refractory Acute Leukemia Based on Chemosensitivity and Genomics/Gene Expression Data</p>	<p><b>N/A</b></p> <p><b>TARGETS</b> Multiple</p>
<p><b>LOCATIONS:</b> Washington</p>	

TRF# XXXXXXXX

**APPENDIX**

Variants of Unknown Significance

**NOTE** One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

**CHEK2**  
N186S and R145W

**CSF1R**  
E920D

**GATA2**  
P161A

**KDM2B**  
E928V

**NTRK2**  
S167Y

**PC**  
A615S

**RAD50**  
D855Y

SAMPLE

TRF# XXXXXXXX

**APPENDIX**

Genes assayed in FoundationOne®Heme

FoundationOne Heme is designed to include genes known to be somatically altered in human hematologic malignancies, sarcomas, and pediatric cancers that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay utilizes DNA sequencing to interrogate 406 genes as well as selected introns of 31 genes involved in rearrangements, in addition to RNA sequencing of 265 genes. The assay will be updated periodically to reflect new knowledge about cancer biology.

**HEMATOLOGICAL MALIGNANCY DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS**

ABL1	ACTB	AKT1	AKT2	AKT3	ALK	AMER1 (FAM123B or WTX)	APC
APH1A	AR	ARAF	ARFRP1	ARHGAP26 (GRAF)	ARID1A	ARID2	ASXL1
ATM	ATR	ATRX	AURKA	AURKB	AXIN1	AXL	B2M
BARD1	BCL10	BCL11B	BCL2	BCL2L2	BCL6	BCL7A	BCOR
BIRC3	BLM	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BRSK1
BTK	BTLA	C11orf30 (EMSY)	CAD	CALR*	CARD11	CBFB	CBL
CCND2	CCND3	CCNE1	CCT6B	CD22	CD274 (PD-L1)	CD36	CD58
CD79A	CD79B	CDC73	CDH1	CDK12	CDK4	CDK6	CDK8
CDKN2A	CDKN2B	CDKN2C	CEBPA	CHD2	CHEK1	CHEK2	CIC
CKS1B	CPS1	CREBBP	CRKL	CRLF2	CSF1R	CSF3R	CTCF
CTNNB1	CUX1	CXCR4	DAXX	DDR2	DDX3X	DNM2	DNMT3A
DTX1	DUSP2	DUSP9	EBF1	ECT2L	EED	EGFR	ELP2
EPHA3	EPHA5	EPHA7	EPHB1	ERBB2	ERBB3	ERBB4	ERG
ETS1	ETV6	EXOSC6	EZH2	FAF1	FAM46C	FANCA	FANCC
FANCE	FANCF	FANCG	FANCL	FAS (TNFRSF6)	FBXO11	FBXO31	FBXW7
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2
FGFR4	FHIT	FLCN	FLT1	FLT3	FLT4	FLYWCH1	FOXL2
FOXO3	FOXP1	FRS2	GADD45B	GATA1	GATA2	GATA3	GID4 (C17orf39)
GNA12	GNA13	GNAQ	GNAS	GPR124	GRIN2A	GSK3B	GTSE1
HDAC4	HDAC7	HGF	HIST1H1C	HIST1H1D	HIST1H1E	HIST1H2AC	HIST1H2AG
HIST1H2AM	HIST1H2BC	HIST1H2BJ	HIST1H2BK	HIST1H2BO	HIST1H3B	HNF1A	HRAS
ICK	ID3	IDH1	IDH2	IGF1R	IKBKE	IKZF1	IKZF2
IL7R	INHBA	INPP4B	INPP5D (SHIP)	IRF1	IRF4	IRF8	IRS2
JAK2	JAK3	JARID2	JUN	KAT6A (MYST3)	KDM2B	KDM4C	KDM5A
KDM6A	KDR	KEAP1	KIT	KLHL6	KMT2A (MLL)	KMT2C (MLL3)	KMT2D (MLL2)
LEF1	LRP1B	LRRK2	MAF	MAFB	MAGED1	MALT1	MAP2K1
MAP2K4	MAP3K1	MAP3K14	MAP3K6	MAP3K7	MAPK1	MCL1	MDM2
MED12	MEF2B	MEF2C	MEN1	MET	MIB1	MITF	MKI67
MPL	MRE11A	MSH2	MSH3	MSH6	MTOR	MUTYH	MYC
MYCN	MYD88	MYO18A	NCOR2	NCSTN	NF1	NF2	NFE2L2
NKX2-1	NOD1	NOTCH1	NOTCH2	NPM1	NRAS	NT5C2	NTRK1
NTRK3	NUP93	NUP98	P2RY8	PAG1	PAK3	PALB2	PASK
PBRM1	PC	PCBP1	PCLO	PDCD1	PDCD11	PDCD1LG2 (PD-L2)	PDGFRA
PDK1	PHF6	PIK3CA	PIK3CG	PIK3R1	PIK3R2	PIM1	PLCG2
PPP2R1A	PRDM1	PRKAR1A	PRKDC	PRSS8	PTCH1	PTEN	PTPN11
PTPN6 (SHP-1)	PTPRO	RAD21	RAD50	RAD51	RAF1	RARA	RASGEF1A
RELN	RET	RHOA	RICTOR	RNF43	ROS1	RPTOR	RUNX1
SDHA	SDHB	SDHC	SDHD	SERP2	SETBP1	SETD2	SF3B1
SMAD2	SMAD4	SMARCA1	SMARCA4	SMARCB1	SMC1A	SMC3	SMO
SOC32	SOC33	SOX10	SOX2	SPEN	SPOP	SRC	SRSF2
STAT3	STAT4	STAT5A	STAT5B	STAT6	STK11	SUFU	SUZ12
TBL1XR1	TCF3 (E2A)	TCL1A (TCL1)	TET2	TGFBR2	TLL2	TMEM30A	TMSB4XP8 (TMSL3)
TNFAIP3	TNFRSF11A	TNFRSF14	TNFRSF17	TOP1	TP53	TP63	TRAF2
TRAF5	TSC1	TSC2	TSHR	TUSC3	TYK2	U2AF1	U2AF2
WDR90	WHSC1 (MMSET or NSD2)	WISP3	WT1	XBP1	XPO1	YY1AP1	ZMYM3
ZNF217	ZNF24 (ZSCAN3)	ZNF703	ZRSR2				

\* Note: the assay was updated on 11/8/2016 to include the detection of alterations in CALR

TRF# XXXXXXXX

**APPENDIX**

Genes assayed in FoundationOne®Heme

**HEMATOLOGICAL MALIGNANCY DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS**

ALK	BCL2	BCL6	BCR	BRAF	CCND1	IGH	IGK	CRLF2
EGFR	EPOR	ETV1	ETV4	ETV5	ETV6	EWSR1	FGFR2	IGL
JAK1	JAK2	KMT2A (MLL)	MYC	NTRK1	PDGFRA	PDGFRB	RAF1	RARA
RET	ROS1	TMPRSS2	TRG					

**HEMATOLOGICAL MALIGNANCY RNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS**

ABI1	ABL1	ABL2	ACSL6	AFF1	AFF4	ALK	ARHGAP26 (GRAF)	
ARHGEF12	ARID1A	ARNT	ASXL1	ATF1	ATG5	ATIC	BCL10	BCL11A
BCL11B	BCL2	BCL3	BCL6	BCL7A	BCL9	BCOR	BCR	BIRC3
BRAF	BTG1	CAMTA1	CARS	CBFA2T3	CBFB	CBL	CCND1	CCND2
CCND3	CD274 (PD-L1)	CDK6	CDX2	CHIC2	CHN1	CIC	CIITA	CLP1
CLTC	CLTCL1	CNTRL (CEP110)	COL1A1	CREB3L1	CREB3L2	CREBBP	CRLF2	CSF1
CTNNB1	DDIT3	DDX10	DDX6	DEK	DUSP22	EGFR	EIF4A2	ELF4
ELL	ELN	EML4	EP300	EPOR	EPS15	ERBB2	ERG	ETS1
ETV1	ETV4	ETV5	ETV6	EWSR1	FCGR2B	FCRL4	FEV	FGFR1
FGFR10P	FGFR2	FGFR3	FLI1	FNBP1	FOXO1	FOXO3	FOXO4	FOXP1
FSTL3	FUS	GAS7	GLI1	GMPS	GPHN	HERPUD1	HEY1	HIP1
HIST1H4I	HLF	HMGA1	HMGA2	HOXA11	HOXA13	HOXA3	HOXA9	HOXC11
HOXC13	HOXD11	HOXD13	HSP90AA1	HSP90AB1	IGH	IGK	IGL	IKZF1
IL21R	IL3	IRF4	ITK	JAK1	JAK2	JAK3	JAZF1	KAT6A (MYST3)
KDSR	KIF5B	KMT2A (MLL)	LASP1	LCP1	LMO1	LMO2	LPP	LYL1
MAF	MAFB	MALT1	MDS2	MECOM	MKL1	MLF1	MLLT1 (ENL)	MLLT10 (AF10)
MLLT3	MLLT4 (AF6)	MLLT6	MN1	MXN1	MSI2	MSN	MUC1	MYB
MYC	MYH11	MYH9	NACA	NBEAP1 (BCL8)	NCOA2	NDRG1	NF1	NF2
NFKB2	NIN	NOTCH1	NPM1	NR4A3	NSD1	NTRK1	NTRK2	NTRK3
NUMA1	NUP214	NUP98	NUTM2A	OMD	P2RY8	PAFAH1B2	PAX3	PAX5
PAX7	PBX1	PCM1	PCSK7	PDCD1LG2 (PD-L2)	PDE4DIP	PDGFB	PDGFRA	PDGFRB
PER1	PHF1	PICALM	PIM1	PLAG1	PML	POU2AF1	PPP1CB	PRDM1
PRDM16	PRRX1	PSIP1	PTCH1	PTK7	RABEP1	RAF1	RALGDS	RAP1GDS1
RARA	RBM15	RET	RHOH	RNF213	ROS1	RPL22	RPN1	RUNX1
RUNX1T1 (ETO)	RUNX2	SEC31A	SEPT5	SEPT6	SEPT9	SET	SH3GL1	SLC1A2
SNX29 (RUNDC2A)	SRSF3	SS18	SSX1	SSX2	SSX4	STAT6	STL	SYK
TAF15	TAL1	TAL2	TBL1XR1	TCF3 (E2A)	TCL1A (TCL1)	TEC	TET1	TFE3
TFG	TFPT	TFRC	TLX1	TLX3	TMPRSS2	TNFRSF11A	TOP1	TP63
TPM3	TPM4	TRIM24	TRIP11	TTL	TYK2	USP6	WHSC1 (MMSET or NSD2)	
WHSC1L1	YPEL5	ZBTB16	ZMYM2	ZNF384	ZNF521			

**ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS**

 Microsatellite (MS) status  
 Tumor Mutational Burden (TMB)



TRF# XXXXXXXX

**APPENDIX**

## Performance Specifications

The median exon coverage for this sample is 843X

**ACCURACY**

Sensitivity: Base Substitutions	At ≥5% Minor Allele Frequency	>99.0%
Sensitivity: Insertions/Deletions (1-40bp)	At ≥10% Minor Allele Frequency	98.0%
Sensitivity: Focal Copy Number Alterations (Homozygous Deletions or Amplifications)	At ≥8% copies	>95.0%
Sensitivity: Microsatellite status	At ≥20% tumor nuclei	97.0%
Sensitivity: Known Gene Fusions	>95.0%	
Specificity: Base Substitutions, Insertions/Deletions, and Focal Copy Number Alterations	Positive Predictive Value (PPV)	>99.0%
Specificity: Known Gene Fusions	Positive Predictive Value (PPV)	>95.0%
Specificity: Microsatellite status	Positive Predictive Value (PPV)	>95.0%
Accuracy: Tumor Mutational Burden	At ≥20% tumor nuclei	>90.0%
Reproducibility (average concordance between replicates)	97.0% inter-batch precision 97.0% intra-batch precision 95.0% microsatellite status precision 96.0% tumor mutation burden precision	

Assay specifications were determined for typical median exon coverage of approximately 500X. For additional information regarding the validation of FoundationOne, please refer to the article, Frampton, GM, et al. Development and validation of a clinical cancer genomic profiling test based on massively parallel DNA sequencing, Nat Biotechnol (2013 Oct. 20).

Microsatellite status, which is a measure of microsatellite instability (MSI), is determined by assessing indel characteristics at 114 homopolymer repeat loci in or near the targeted gene regions of the FoundationOne Heme test. Microsatellite status is assayed for all FoundationOne Heme samples and may be reported as "MSI-High", "MSI-Intermediate", "MS-Stable", or "Cannot Be Determined". Microsatellite status is reported as "Cannot Be Determined" if the sample is not of sufficient quality to be confidently determined.

Tumor Mutational Burden (TMB) is determined by measuring the number of somatic mutations occurring in sequenced genes on the FoundationOne and FoundationOne Heme tests and extrapolating to the genome as a whole. TMB is assayed for all FoundationOne and FoundationOne Heme samples and may be reported as "TMB-High", "TMB-Intermediate", "TMB-Low", or "Cannot Be Determined". TMB results, which are rounded to the nearest integer, are determined as follows: TMB-High corresponds to greater than or equal to 20 mutations per megabase (Muts/Mb); TMB-Intermediate corresponds to 6-19 Muts/Mb; TMB-Low corresponds to fewer than or equal to 5 Muts/Mb. Tumor Mutational Burden is reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine Tumor Mutational Burden.

For additional information specific to the performance of this specimen, please contact Foundation Medicine, Inc. at 1-888-988-3639.

TRF# XXXXXXXX

**APPENDIX**

About FoundationOne®Heme

## ABOUT FOUNDATIONONE HEME

FoundationOne Heme is a comprehensive genomic profiling test for hematologic malignancies, sarcomas and pediatric cancers. The test is designed to provide physicians with clinically actionable information to help with diagnostic sub-classification, prognosis assessment, and targeted therapeutic selection. Test results provide information about clinically significant alterations, potential targeted therapies, available clinical trials, and quantitative markers that may support immunotherapy clinical trial enrollment. FoundationOne Heme is analytically validated to detect all classes of genomic alterations in more than 400 cancer-related genes. In addition to DNA sequencing, FoundationOne Heme employs RNA sequencing across more than 250 genes to capture a broad range of gene fusions, common drivers of hematologic malignancies and sarcomas, pediatric cancers.

FoundationOne Heme was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne has not been cleared or approved by the United States Food and Drug Administration (FDA). The FDA has determined that such clearance or approval is not necessary. FoundationOne Heme may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratory is certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA) as qualified to perform high-complexity clinical testing.

## THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. *Note:* A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

### Diagnostic Significance

FoundationOne Heme identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

### Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification – equivocal" implies that FoundationOne Heme data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne Heme for identifying a copy number amplification is five (5) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that FoundationOne Heme data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that FoundationOne Heme analytical methodology has identified as being present in <10% of the assayed tumor DNA.

### Ranking of Alterations and Therapies

#### Biomarker Findings

Appear at the top of the report, but are not ranked higher than Genomic Findings.

#### Genomic Findings

Therapies with Clinical Benefit In Patient's Tumor Type → Therapies with Clinical Benefit in Other Tumor Type → Clinical Trial Options → No Known Options (If multiple findings exist within any of these categories, the results are listed alphabetically by gene name.)

#### Therapies

Sensitizing therapies → Resistant therapies. (If multiple therapies exist within any of these categories, they are listed alphabetically by therapy name.)

#### Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

### LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

### NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

### NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or

governmental, will reimburse a patient for the cost of FoundationOne Heme.

## TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. These include: subclonal alterations in heterogeneous samples, low sample quality or with homozygous losses of <3 exons; and deletions and insertions >40bp, or in repetitive/high homology sequences. FoundationOne Heme is performed using DNA and RNA derived from tumor, and as such germline events may not be reported.

The following targets typically have low coverage resulting in a reduction in sensitivity: SDHD exon 4, TNFRSF11A exon1, and TP53 exon 1.

FoundationOne Heme complies with all European Union (EU) requirements of the IVD Directive 98/79EC. As such, the FoundationOne Heme Assay has been registered for CE mark by our EU Authorized Representative, Qarad b.v.b.a, Cijpestraat 3, 2440 Geel, Belgium.



TRF# XXXXXXXX

**APPENDIX**

About FoundationOne®Heme

**SELECT ABBREVIATIONS**

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

SAMPLE

TRF# XXXXXXXX

**APPENDIX**      **References**

1. Linnekin D (1999) Early signaling pathways activated by c-Kit in hematopoietic cells. *Int J Biochem Cell Biol* 31(10):1053-74.
2. Fletcher JA (2004) Role of KIT and platelet-derived growth factor receptors as oncoproteins. *Semin Oncol* 31(2 Suppl 6): 4-11.
3. Agarwal R, Robson M (2009) Inherited predisposition to gastrointestinal stromal tumor. *Hematol Oncol Clin North Am* 23(1):1-13, vii.
4. Miettinen M, Lasota J (2006) Gastrointestinal stromal tumors: review on morphology, molecular pathology, prognosis, and differential diagnosis. *Arch Pathol Lab Med* 130(10):1466-78.
5. Guo T, Hajdu M, Agaram NP, et al. (2009) Mechanisms of sunitinib resistance in gastrointestinal stromal tumors harboring KITAY502-3ins mutation: an in vitro mutagenesis screen for drug resistance. *Clin Cancer Res* 15(22):6862-70.
6. Zhang HM, Yu X, Greig MJ, et al. (2010) Drug binding and resistance mechanism of KIT tyrosine kinase revealed by hydrogen/deuterium exchange FTICR mass spectrometry. *Protein Sci* 19(4):703-15.
7. Ma Y, Zeng S, Metcalfe DD, et al. (2002) The c-KIT mutation causing human mastocytosis is resistant to ST1571 and other KIT kinase inhibitors; kinases with enzymatic site mutations show different inhibitor sensitivity profiles than wild-type kinases and those with regulatory-type mutations. *Blood* 99(5):1741-4.
8. Italiano A, Sirvent N, Michiels JF, et al. (2005) Tumour response to paclitaxel in an adult with relapsed nephroblastoma. *Lancet Oncol* 6(4):252-3.
9. Wardelmann E, Merkelbach-Bruse S, Pauls K, et al. (2006) Polyclonal evolution of multiple secondary KIT mutations in gastrointestinal stromal tumors under treatment with imatinib mesylate. *Clin Cancer Res* 12(6):1743-9.
10. Heinrich MC, Maki RG, Corless CL, et al. (2008) Primary and secondary kinase genotypes correlate with the biological and clinical activity of sunitinib in imatinib-resistant gastrointestinal stromal tumor. *J Clin Oncol* 26(33):5352-9.
11. Heinrich MC, Corless CL, Blanke CD, et al. (2006) Molecular correlates of imatinib resistance in gastrointestinal stromal tumors. *J Clin Oncol* 24(29):4764-74.
12. Vega-Ruiz A, Cortes JE, Sever M, et al. (2009) Phase II study of imatinib mesylate as therapy for patients with systemic mastocytosis. *Leuk Res* 33(11):1481-4.
13. Antonescu CR, Besmer P, Guo T, et al. (2005) Acquired resistance to imatinib in gastrointestinal stromal tumor occurs through secondary gene mutation. *Clin Cancer Res* 11(11): 4182-90.
14. Liegl B, Kepten I, Le C, et al. (2008) Heterogeneity of kinase inhibitor resistance mechanisms in GIST. *J Pathol* 216(1):64-74.
15. Zermati Y, De Sepulveda P, Féger F, et al. (2003) Effect of tyrosine kinase inhibitor ST1571 on the kinase activity of wild-type and various mutated c-kit receptors found in mast cell neoplasms. *Oncogene* 22(5):660-4.
16. Droogendijk HJ, Kluin-Nelemans HJ, van Doornaal JJ, et al. (2006) Imatinib mesylate in the treatment of systemic mastocytosis: a phase II trial. *Cancer* 107(2):345-51.
17. Garner AP, Gozgit JM, Anjum R, et al. (2014) Ponatinib inhibits polyclonal drug-resistant KIT oncoproteins and shows therapeutic potential in heavily pretreated gastrointestinal stromal tumor (GIST) patients. *Clin Cancer Res* ePub Sep 2014.
18. Gajjala KS, Wu JC, Christensen J, et al. (2009) KIT kinase mutants show unique mechanisms of drug resistance to imatinib and sunitinib in gastrointestinal stromal tumor patients. *Proc Natl Acad Sci USA* 106(5):1542-7.
19. Dziadziuszko R, Le AT, Wrona A, et al. (2016) An Activating KIT Mutation Induces Crizotinib Resistance in ROS1-Positive Lung Cancer. *J Thorac Oncol* ePub Apr 2016.
20. Debiec-Rychter M, Sciort R, Le Cesne A, et al. (2006) KIT mutations and dose selection for imatinib in patients with advanced gastrointestinal stromal tumours. *Eur J Cancer* 42(8): 1093-103.
21. Evans EK, Gardino AK, Kim JL, et al. (2017) A precision therapy against cancers driven by KIT/PDGFR mutations. *Sci Transl Med* 9(414).
22. Cancer Genome Atlas Research Network (2013) Genomic and epigenomic landscapes of adult de novo acute myeloid leukemia. *N Engl J Med* 368(22):2059-74.
23. Malaise M, Steinbach D, Corbacioglu S (2009) Clinical implications of c-KIT mutations in acute myelogenous leukemia. *Curr Hematol Malig Rep* 4(2):77-82.
24. Paschka P, Döhner K (2013) Core-binding factor acute myeloid leukemia: can we improve on HiDAC consolidation? *Hematology Am Soc Hematol Educ Program* 2013:209-19.
25. Riera L, Marmont F, Toppino D, et al. (2013) Core binding factor acute myeloid leukaemia and c-KIT mutations. *Oncol Rep* 29(5):1867-72.
26. Manara E, Bisio V, Masetti R, et al. (2014) Core-binding factor acute myeloid leukemia in pediatric patients enrolled in the AIEOP AML 2002/01 trial: screening and prognostic impact of c-KIT mutations. *Leukemia* 28(5):1132-4.
27. Marcucci G, Haferlach T, Döhner H (2011) Molecular genetics of adult acute myeloid leukemia: prognostic and therapeutic implications. *J Clin Oncol* 29(5):475-86.
28. Abbaspour Babaei M, Kamalidehghan B, Saleem M, et al. (2016) Receptor tyrosine kinase (c-Kit) inhibitors: a potential therapeutic target in cancer cells. *Drug Des Devel Ther* 10:2443-59.
29. Ramaswamy A, Pande N, Shetty O, et al. (2016) Pazopanib in metastatic multiply treated progressive gastrointestinal stromal tumors: feasible and efficacious. *J Gastrointest Oncol* 7(4):638-43.
30. Demetri GD, Reichardt P, Kang YK, et al. (2013) Efficacy and safety of regorafenib for advanced gastrointestinal stromal tumours after failure of imatinib and sunitinib (GRID): an international, multicentre, randomised, placebo-controlled, phase 3 trial. *Lancet* 381(9863):295-302.
31. Gotlib J, Kluin-Nelemans HC, George TI, et al. (2016) Efficacy and Safety of Midostaurin in Advanced Systemic Mastocytosis. *N Engl J Med* 374(26):2530-41.
32. Jawhar M, Schwaab J, Naumann N, et al. (2017) Response and progression on midostaurin in advanced systemic mastocytosis: KIT D816V and other molecular markers. *Blood* ePub Apr 2017.
33. Xu X, Kreisel FH, Frater JL, et al. (2014) Mast cell leukemia with prolonged survival on PKC412/midostaurin. *Int J Clin Exp Pathol* 7(6):3439-43.
34. Gotlib J, Berubé C, Growney JD, et al. (2005) Activity of the tyrosine kinase inhibitor PKC412 in a patient with mast cell leukemia with the D816V KIT mutation. *Blood* 106(8):2865-70.
35. Gao J, Tian Y, Li J, et al. (2013) Secondary mutations of c-KIT contribute to acquired resistance to imatinib and decrease efficacy of sunitinib in Chinese patients with gastrointestinal stromal tumors. *Med Oncol* 30(2):522.
36. Heydt C, Kumm N, Fassunke J, et al. (2015) Massively parallel sequencing fails to detect minor resistant subclones in tissue samples prior to tyrosine kinase inhibitor therapy. *BMC Cancer* 15:291.
37. Nishida T, Takahashi T, Nishitani A, et al. (2009) Sunitinib-resistant gastrointestinal stromal tumors harbor cis-mutations in the activation loop of the KIT gene. *Int J Clin Oncol* 14(2): 143-9.
38. Guo T, Agaram NP, Wong GC, et al. (2007) Sorafenib inhibits the imatinib-resistant KITT670I gatekeeper mutation in gastrointestinal stromal tumor. *Clin Cancer Res* 13(16):4874-81.
39. Heinrich MC, Marino-Enriquez A, Presnell A, et al. (2012) Sorafenib inhibits many kinase mutations associated with drug-resistant gastrointestinal stromal tumors. *Mol Cancer Ther* 11(8):1770-80.
40. Hu S, Niu H, Minkin P, et al. (2008) Comparison of antitumor effects of multitargeted tyrosine kinase inhibitors in acute myelogenous leukemia. *Mol Cancer Ther* 7(5):1110-20.
41. Bisagni G, Rossi G, Cavazza A, et al. (2009) Long lasting response to the multitargeted inhibitor bay 43-9006 (Sorafenib) in a heavily pretreated metastatic thymic carcinoma. *J Thorac Oncol* 4(6):773-5.
42. Handolias D, Hamilton AL, Salemi R, et al. (2010) Clinical responses observed with imatinib or sorafenib in melanoma patients expressing mutations in KIT. *Br J Cancer* 102(8): 1219-23.
43. Singeltary B, Ghose A, Sussman J, et al. (2014) Durable response with a combination of imatinib and sorafenib in KIT exon 17 mutant gastrointestinal stromal tumor. *J Gastrointest Oncol* 5(1):E27-9.
44. Guida T, Anaganti S, Provitera L, et al. (2007) Sorafenib inhibits imatinib-resistant KIT and platelet-derived growth factor receptor beta gatekeeper mutants. *Clin Cancer Res* 13(11): 3363-9.
45. Lierman E, Lahortiga I, Van Miegroet H, et al. (2007) The ability of sorafenib to inhibit oncogenic PDGFRbeta and FLT3 mutants and overcome resistance to other small molecule inhibitors. *Haematologica* 92(1):27-34.
46. Lierman E, Smits S, Cools J, et al. (2012) Ponatinib is active against imatinib-resistant mutants of FIP1L1-PDGFRa and KIT, and against FGFR1-derived fusion kinases. *Leukemia* 26(7): 1693-5.
47. Jin B, Ding K, Pan J (2014) Ponatinib induces apoptosis in imatinib-resistant human mast cells by dephosphorylating mutant D816V KIT and silencing  $\beta$ -catenin signaling. *Mol Cancer Ther* 13(5):1217-30.
48. Gleixner KV, Peter B, Blatt K, et al. (2013) Synergistic growth-inhibitory effects of ponatinib and midostaurin (PKC412) on neoplastic mast cells carrying KIT D816V. *Haematologica* 98(9):1450-7.
49. Gozgit JM, Wong MJ, Wardwell S, et al. (2011) Potent activity of ponatinib (AP24534) in models of FLT3-driven acute myeloid leukemia and other hematologic malignancies. *Mol Cancer Ther* 10(6):1028-35.
50. Si L, Xu X, Kong Y, et al. (2012) Major response to everolimus in melanoma with acquired imatinib resistance. *J Clin Oncol* 30(4):e37-40.
51. Pylayeva-Gupta Y, Grabocka E, Bar-Sagi D (2011) RAS oncogenes: weaving a tumorigenic web. *Nat Rev Cancer* 11(11): 761-74.
52. Buhrman G, Holzapfel G, Fetics S, et al. (2010) Allosteric modulation of Ras positions Q61 for a direct role in catalysis. *Proc Natl Acad Sci USA* 107(11):4931-6.
53. Cirstea IC, Kutsche K, Dvorsky R, et al. (2010) A restricted spectrum of NRAS mutations causes Noonan syndrome. *Nat Genet* 42(1):27-9.
54. Colicelli J (2004) Human RAS superfamily proteins and related GTPases. *Sci STKE* 2004(250):RE13.
55. Edkins S, O'Meara S, Parker A, et al. (2006) Recurrent KRAS codon 146 mutations in human colorectal cancer. *Cancer Biol Ther* 5(8):928-32.
56. Feig LA, Cooper GM (1988) Relationship among guanine nucleotide exchange, GTP hydrolysis, and transforming potential of mutated ras proteins. *Mol Cell Biol* 8(6):2472-8.
57. Janakiraman M, Vakiani E, Zeng Z, et al. (2010) Genomic and biological characterization of exon 4 KRAS mutations in human cancer. *Cancer Res* 70(14):5901-11.
58. Lukman S, Grant BJ, Gorfe AA, et al. (2010) The distinct conformational dynamics of K-Ras and H-Ras A59G. *PLoS Comput Biol* 6(9).
59. Prior IA, Lewis PD, Mattos C (2012) A comprehensive survey of Ras mutations in cancer. *Cancer Res* 72(10):2457-67.
60. Privé GG, Milburn MV, Tong L, et al. (1992) X-ray crystal structures of transforming p21 ras mutants suggest a transition-state stabilization mechanism for GTP hydrolysis. *Proc Natl Acad Sci USA* 89(8):3649-53.
61. Runtuwene V, van Eekelen M, Overvoorde J, et al. (2011) Noonan syndrome gain-of-function mutations in NRAS cause zebrafish gastrulation defects. *Dis Model Mech* 4(3):393-9.
62. Scheffzek K, Ahmadian MR, Kabsch W, et al. (1997) The Ras-RasGAP complex: structural basis for GTPase activation and its loss in oncogenic Ras mutants. *Science* 277(5324):333-8.